

Mast cells granular contents are crucial for deep vein thrombosis in mice

Ponomaryov, Tatyana; Payne, Holly; Fabritz, Larissa; Wagner, Denisa D; Brill, Alexander

DOI:

[10.1161/CIRCRESAHA.117.311185](https://doi.org/10.1161/CIRCRESAHA.117.311185)

License:

Creative Commons: Attribution (CC BY)

Document Version

Publisher's PDF, also known as Version of record

Citation for published version (Harvard):

Ponomaryov, T, Payne, H, Fabritz, L, Wagner, DD & Brill, A 2017, 'Mast cells granular contents are crucial for deep vein thrombosis in mice', *Circulation Research*, vol. 121, no. 8, pp. 941-950.
<https://doi.org/10.1161/CIRCRESAHA.117.311185>

[Link to publication on Research at Birmingham portal](#)

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Mast Cells Granular Contents Are Crucial for Deep Vein Thrombosis in Mice

Tatyana Ponomaryov,* Holly Payne,* Larissa Fabritz, Denisa D. Wagner, Alexander Brill

Rationale: Deep vein thrombosis (DVT) and its complication pulmonary embolism have high morbidity reducing quality of life and leading to death. Cellular mechanisms of DVT initiation remain poorly understood.

Objective: We sought to determine the role of mast cells (MCs) in DVT initiation and validate MCs as a potential target for DVT prevention.

Methods and Results: In a mouse model, DVT was induced by partial ligation (stenosis) of the inferior vena cava. We demonstrated that 2 strains of mice deficient for MCs were completely protected from DVT. Adoptive transfer of in vitro differentiated MCs restored thrombosis. MCs were present in the venous wall, and the number of granule-containing MCs decreased with thrombosis. Pharmacological depletion of MCs granules or prevention of MC degranulation also reduced DVT. Basal plasma levels of von Willebrand factor and recruitment of platelets to the inferior vena cava wall after DVT induction were reduced in MC-deficient mice. Stenosis application increased plasma levels of soluble P-selectin in wild-type but not in MC-deficient mice. MC releasate elevated ICAM-1 (intercellular adhesion molecule-1) expression on HUVEC (human umbilical vein endothelial cells) in vitro. Topical application of compound 48/80, an MC secretagogue, or histamine, a Weibel–Palade body secretagogue from MCs, potentiated DVT in wild-type mice, and histamine restored thrombosis in MC-deficient animals.

Conclusions: MCs exacerbate DVT likely through endothelial activation and Weibel–Palade body release, which is, at least in part, mediated by histamine. Because MCs do not directly contribute to normal hemostasis, they can be considered potential targets for prevention of DVT in humans. (*Circ Res.* 2017;121:941-950. DOI: 10.1161/CIRCRESAHA.117.311185.)

Key Words: endothelium ■ histamine ■ mast cells ■ venous thrombosis

Venous thromboembolism, designating deep vein thrombosis (DVT) together with pulmonary embolism (PE), is a severe disease frequently diagnosed in the Western world. This pathological condition develops in ≈100 to 200 per 100 000 individuals annually.¹ In a part of patients (≈69 per 100 000),² DVT is complicated by PE, in which blood clot detaches and reaches lungs, leading to death in ≈30% of cases.³ As per estimation of the American Surgeon General, these diseases are responsible for >100 000 deaths a year in the United States only,⁴ exceeding mortality after myocardium infarction or stroke.⁵ Even after successful treatment, DVT and PE may result in post-thrombotic syndrome and chronic thromboembolic pulmonary hypertension, respectively,^{6,7} causing poor life quality and requiring additional medical attention, including prolonged period of anticoagulation therapy.

Editorial, see p 899
Meet the First Author, see p 894

Mechanisms of thrombus development in veins remain incompletely understood. Risk factors for DVT include those that cannot be avoided or changed (such as old age or cancer) and various medical conditions, such as obesity, hypertension, and atrial fibrillation.^{8,9} About 60% of patients, who develop DVT and PE after hospitalization or surgery, develop the disease within 90 days,⁸ demonstrating connection between previous invasive therapy and DVT formation.

From the pathogenetic point of view, factors triggering thrombosis in veins differ from those in arteries. Arterial thrombosis usually starts with atherosclerotic plaque rupture with exposure of the subendothelial adhesive proteins, to which platelets bind forming

Original received April 16, 2017; revision received July 18, 2017; accepted July 21, 2017. In June 2017, the average time from submission to first decision for all original research papers submitted to *Circulation Research* was 12.45 days.

From the Institute of Cardiovascular Sciences, College of Medical and Dental Sciences, University of Birmingham, United Kingdom (T.P., H.P., L.F., A.B.); Department of Cardiology, University Hospital Birmingham, United Kingdom (L.F.); Program in Cellular and Molecular Medicine (D.D.W., A.B.) and Division of Hematology/Oncology (D.D.W., A.B.), Boston Children's Hospital, MA; and Department of Pediatrics, Harvard Medical School, Boston, MA (D.D.W., A.B.).

*T.P. and H.P. contributed equally to this article.

The online-only Data Supplement is available with this article at <http://circres.ahajournals.org/lookup/suppl/doi:10.1161/CIRCRESAHA.117.311185/-/DC1>.

Correspondence to Alexander Brill, MD, PhD, Institute of Cardiovascular Sciences, College of Medical and Dental Sciences, University of Birmingham, Edgbaston, Birmingham B15 2TT, United Kingdom. E-mail a.brill@bham.ac.uk

© 2017 The Authors. *Circulation Research* is published on behalf of the American Heart Association, Inc., by Wolters Kluwer Health, Inc. This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution, and reproduction in any medium, provided that the original work is properly cited.

Circulation Research is available at <http://circres.ahajournals.org>

DOI: 10.1161/CIRCRESAHA.117.311185

Novelty and Significance

What Is Known?

- Development of deep vein thrombosis (DVT) includes mechanisms similar to sterile inflammation.
- Mast cells (MCs) reside in the vicinity to blood vessels.
- MCs contain granular mediators affecting both thrombosis and inflammation.

What New Information Does This Article Contribute?

- MC granular constituents promote DVT in mice.
- Genetic depletion or pharmacological inhibition of MC granule release protects mice from DVT.
- Targeting MCs may be a safe and efficient strategy to prevent DVT.

Current methods of DVT prophylaxis are usually based on inhibition of blood coagulation factors. Because these factors are involved in normal bleeding arrest, such strategy results in bleeding complications in a part of patients, and, therefore, new approaches are required. We demonstrated that MCs induce DVT in a mouse model. Mice lacking MCs were protected against DVT. Pharmacological ablation of MCs or inhibition of their degranulation also prevented venous thrombosis. Local stimulation of MC degranulation or release of Weibel–Palade bodies from endothelial cells strongly promoted DVT. These findings suggest that proinflammatory mediators released from MC granules are able to cause DVT. Given that MC deficiency is not accompanied by impaired hemostasis and therefore unlikely to provoke bleeding complications, MCs may be a target for DVT prophylaxis in humans.

Nonstandard Abbreviations and Acronyms

DVT	deep vein thrombosis
FcεRI	immunoglobulin E receptor I
IVC	inferior vena cava
MC	mast cells
PE	pulmonary embolism
vWF	von Willebrand Factor
WPB	Weibel–Palade body
WT	wild type

a relatively small platelet-rich thrombus. In contrast, in veins, large thrombi enriched by red blood cells develop without signs of endothelial denudation.¹⁰ DVT begins in venous valvular sinus, in which blood flow can become stagnant resulting in prolonged time of blood exchange in valves, or stasis, an element of the Virchow triad.¹¹ In venous valves, hypoxia and elevated hematocrit have been observed, which may increase local thrombogenicity.¹² The role of blood flow stagnancy increases in people in bed-ridden position caused by surgery or other situations accompanied by total or partial immobilization (limb paralysis, long-haul flights, etc).

We and others have demonstrated that an inflammatory component plays a role in triggering DVT.^{13,14} Local hypoxia may result in release of Weibel–Palade body (WPB) constituents, such as von Willebrand factor (vWF) and P-selectin, to the endothelial surface leading to recruitment of platelets and leukocytes.¹⁵ Platelets can contribute to thrombosis initiation likely by providing their procoagulant surface to clotting factors, whereas neutrophils support DVT propagation by producing neutrophil extracellular traps.^{16,17}

Mast cells (MCs) are a part of the innate immune system. They originate from the hematopoietic progenitor in bone marrow and enter the circulation as early lineage progenitors.¹⁸ After egression to tissues and maturation, MCs express a specific set of antigens including FcεRI (a receptor for immunoglobulin E) and Kit (a receptor for stem cell factor). The role of MCs in allergic inflammation, a risk factor for DVT and PE,¹⁹ has been well documented.²⁰ MC granules contain potent anticoagulants, such as heparin and tPA (tissue-type plasminogen activator);

endothelial activators (histamine and tumor necrosis factor-α); and many enzymes (tryptases, chymases, and others).^{21,22} In tissues, MCs are located in the vicinity of blood vessels,²³ and their presence at the site of human DVT has been reported.²⁴

Herein, using murine models, we demonstrate that liberation of MC granule constituents is critical for DVT initiation. This is a novel mechanism regulating venous thrombosis, which could potentially be used to target DVT in humans.

Methods

Animals

MC-deficient Kit^{W^v} (Jackson Laboratory 000049, backcrossed to C57BL/6J for decades) mice²⁵ and Kit^{W^{sh}} (Jackson Laboratory 012861, backcrossed to C57BL/6J for 11 generations) mice^{26,27} were purchased in the Jackson Laboratory, and colonies were maintained in animal facilities at Children's Hospital Boston and the University of Birmingham under standard conditions. Wild-type littermates were used as a control in DVT and intravital microscopy experiments. C57BL/6 mice from Charles River were used in experiments with topical application of histamine and compound 48/80. All animal experiments were approved by either Institutional Animal Review Board at Children's Hospital Boston or Animal Welfare Ethical Review Body and the UK Home Office (United Kingdom; Project Licenses 40/3745 and 70/8286).

DVT Surgery

The stenosis of the inferior vena cava (IVC) was performed as described previously.¹³ In brief, mice were anesthetized with isoflurane–oxygen mixture, midabdominal incision was performed, and intestines were exteriorized and soaked in warm saline. All the IVC side branches were ligated. The IVC was gently isolated from aorta, a 7-0 polypropylene suture was placed over the IVC, and ligated over a spacer (30-gauge needle), and then, the spacer was removed. This method produces IVC lumen area reduction by ≈90% and is not accompanied by endothelial denudation. Finally, peritoneum was closed with silk suture and skin with either suture or staples. Thrombosis was evaluated at different time points ≤48 hours, and thrombi were excised for measurement. Assessment of thrombus weight and length by scale and ruler, respectively, has certain limitations; for example, precision of length measurement is ≈0.5 mm, whereas weight determination may vary ±1 to 1.5 mg. However, area of thrombi measured by ultrasonography and thrombus weight determined by scale demonstrate strong correlation ($r^2=0.96$)²⁸; and therefore, the manual method of thrombus size assessment used in this study can be considered valid.

Culture of Murine Bone Marrow–Derived MCs

Bone marrow–derived MC culture was established as described elsewhere.²⁹ In brief, bone marrow cells were flushed from adult wild-type (WT) mice femurs and tibia, centrifuged, and resuspended in RPMI (Roswell Park Memorial Institute) medium containing 10% fetal bovine serum, 100 U/mL penicillin, 0.1 mg/mL streptomycin, 25 mmol/L HEPES, 2 mmol/L L-glutamine, 1 mmol/L sodium pyruvate, 1 mmol/L nonessential amino acids, and 1 mmol/L MEM (minimal essential medium) amino acids (all from Sigma) in the presence of 5 ng/mL recombinant mouse interleukin-3 (Peprotech). A selection process for mature MCs was for a period of at least 4 weeks with continuous enrichment for nonadherent fraction of MC precursors. After this period, cells were phenotypically >90% mature as assessed by fluorescence-activated cell sorter staining with antibodies against mouse FcεR1-PE (eBioscience; clone MAR-1) and CD117(c-Kit)-APC (eBioscience; clone 2B8; Online Figure I). To induce MC degranulation, cells were sensitized overnight with 100 ng/mL mouse anti-DNP-IgE (Sigma), washed, and stimulated with 100 ng/mL DNP-HSA (dinitrophenyl-human serum albumin; Sigma) for 0.5 or 1 hour (fast release) or 6 hours (slow release). The efficiency of mediator release was monitored by β-hexaminidase activity in MC supernatants (releasates; Online Figure I).³⁰

MC Granule Depletion

Mice were pretreated IP every 12 hours for 4 days with the MC-degranulating compound 48/80 dissolved in 200 μL of sterile saline according to the following scheme: first day, 0.6 mg/kg; second day, 1.0 mg/kg; third day, 1.2 mg/kg; fourth day, 2.4 mg/kg. This treatment results in peritoneal MC granule content depletion by ≈84%.³¹ Twenty-four hours after last injection, mice were subjected to IVC stenosis for 48 hours.

MC Membrane Stabilization In Vivo

MC membrane stabilizer sodium cromoglycate (cromolyn, 100 mg/kg body weight; Sigma) or 200 μL of sterile saline was injected IP into mice 24 hours and 30 minutes before surgery. Then, IVC stenosis was applied for 48 hours. Ketotifen (25 mg/kg body weight; Sigma) or 200 μL of sterile saline was injected IP 24 hours and 30 minutes before and 24 hours after IVC stenosis application for 48 hours. Sodium cromoglycate inhibits MC degranulation and has previously been shown to work in mice in the indicated or lower dose.^{32,33}

Murine Doppler Ultrasound

Doppler ultrasound was performed as described previously.^{34,35} Mice were anesthetized with isoflurane 2% plus oxygen, the region of interest was shaved, and covered with ultrasound gel. Images of the vena cava inferior and surrounding structures and vessels were obtained using the Visualsonics Vevo 2100 system with a cardiovascular scanhead.

Determination of Plasma vWF Levels

The test was performed as described previously.¹³ The level of vWF in pooled plasma of 20 C57BL/6 mice served as a control.

Results

MC-Deficient Mice Are Protected Against DVT

To clarify whether MCs play a role in DVT, we subjected mice with W-sh Kit mutation, which lack MCs, to IVC stenosis. Both 24 and 48 hours after stenosis application, none of the mutant mice developed a thrombus compared with 75% and 90% thrombosis in WT controls, respectively (Figure 1A and 1B). To confirm the phenotype, we used another strain of Kit mutants, Kit^{W-v}, with littermate controls. After 48-hour stenosis, 89% of control mice developed a thrombus, whereas no thrombosis was observed in Kit^{W-v} animals (Figure 1C). Further experiments were performed using Kit^{W-sh} mutants with littermate

controls. To verify presence of thrombi in WT and absence in Kit^{W-sh} mice by an independent method, we used ultrasound Doppler (Figure 1D). Control WT mice with a thrombus did not have blood flow in the IVC and their IVC could not be compressed by applying pressure on the mouse abdomen with the probe. In contrast, Kit^{W-sh} mice had clear blood flow in the IVC, and the IVC was easily compressible suggesting absence of thrombi. The protective effect of MC deficiency was specific to DVT because no difference in thrombosis between WT and Kit^{W-sh} mice in ferric chloride–induced thrombosis was observed (Online Figure IIA). This finding indicates that the prothrombotic effect of MCs requires endothelial monolayer, which is denuded by ferric chloride.³⁶ Tail bleeding time and coagulation tests (prothrombin time and activated partial thromboplastin time) in Kit^{W-sh} mice were unchanged compared with WT controls (Online Figure IIB through IID).

MC Reconstitution Restores Thrombosis in MC-Deficient Mice

In addition to the absence of MCs, Kit mutation mediates other abnormalities, such as defective melanogenesis and impaired germ cell development. To rule out any other potential mechanisms, through which mutant Kit could affect thrombosis, we performed adoptive transfer of in vitro differentiated bone marrow MCs to Kit^{W-sh} mice, a procedure that restores MC pool in murine tissues.^{37,38} About 8 to 10 weeks after adoptive transfer, mice were subjected to IVC stenosis. As seen in Figure 1B, 67% of these animals produced a thrombus, which was significantly higher than MC-deficient mice without adoptive transfer and did not differ from thrombosis prevalence in WT controls. Thrombus weight and length in Kit^{W-sh} mice with and without adoptive transfer were also significantly different. Thus, it is the lack of MCs that protects against DVT.

Granule-Containing MCs Are Present in the IVC Wall and Their Number Decreases With Thrombosis

To test MC availability in the IVC, we performed staining of the vessel wall with Toluidine blue, a dye that specifically stains MC granules. MCs were found in the IVC wall in the vicinity to the abluminal side of the endothelium (Figure 2A). Forty-eight hours after stenosis application, the number of MCs in the IVC wall 2 to 3 mm below the ligation decreased in mice that developed thrombi compared with sham-operated animals (12.1±1.0 versus 25.6±2.1; *P*<0.0001; Figure 2B; Online Figure III). This result suggests that in the course of DVT, MCs either migrate out of the vessel wall or degranulate and thus lose Toluidine blue granule staining. The number of MCs in the IVCs of mice after 48-hour stenosis but without thrombi remained unchanged (33.4±8.1; *P*=0.27). This finding implies that degranulation of MCs in the IVC wall is associated with thrombosis.

MC Granule Depletion Prevents DVT

We next assessed whether pharmacological ablation of MCs phenocopy the absence of DVT in Kit^{W-sh} mice. MC granules were depleted by consecutive daily injections of compound 48/80 as described in Methods section. Thrombi developed in 86% of control saline-treated mice, whereas none of the

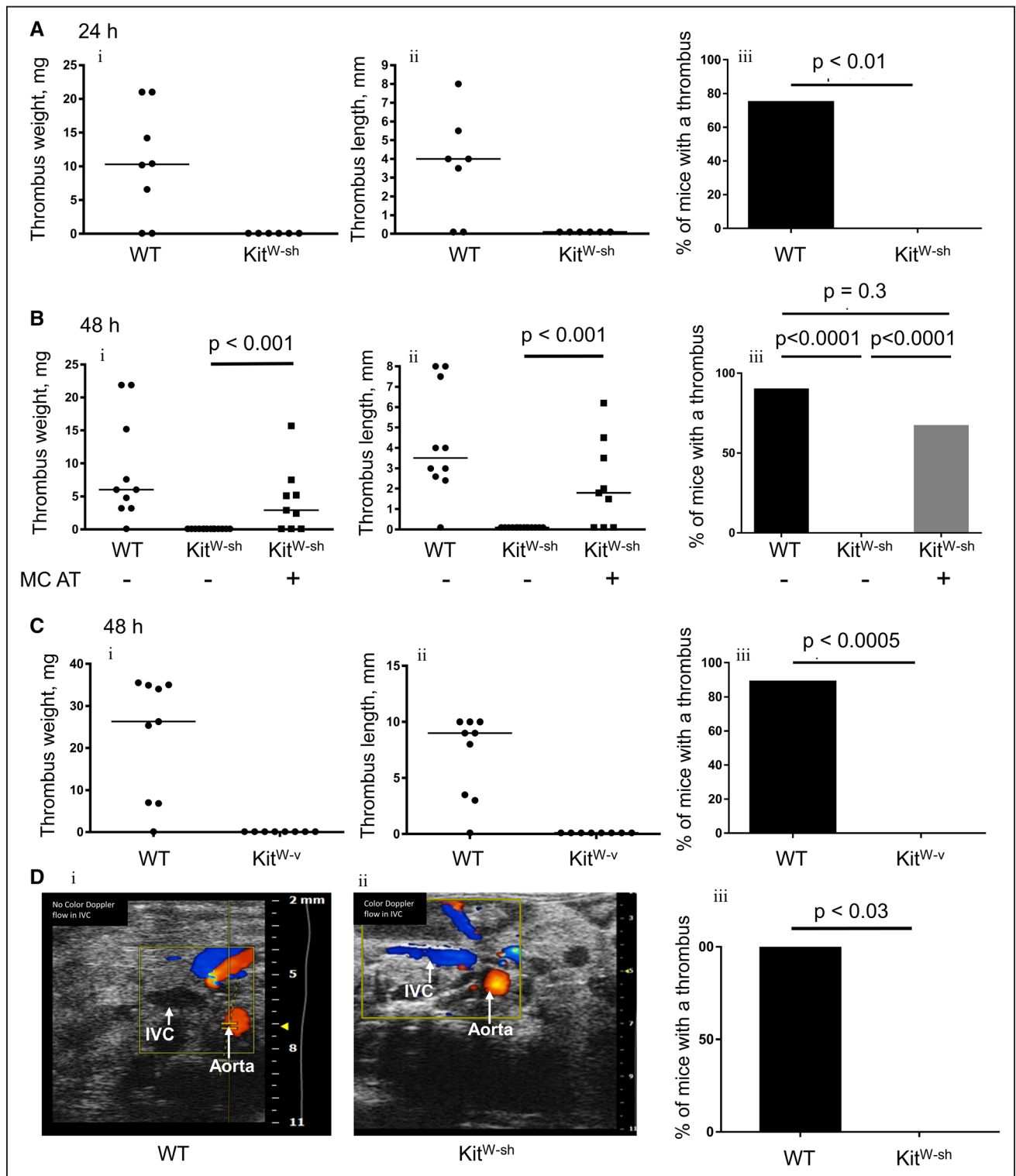


Figure 1. Kit^{W-sh} and Kit^{W-v} mice are protected against deep vein thrombosis, and mast cell (MC) repopulation restores thrombosis. Kit^{W-sh} mice and wild-type (WT) littermates were subjected to 24 h (A) or 48 h (B and C) stenosis of the inferior vena cava (IVC). A separate group of Kit^{W-sh} mice (n=9) had undergone adoptive transfer (AT) of 10×10^6 in vitro differentiated MCs 3 mo before the experiment. A, WT, n=8; Kit^{W-sh} , n=6. B, WT, n=10; Kit^{W-sh} , n=14, $Kit^{W-sh}+AT$, n=9. C, Kit^{W-v} mice (n=8) and WT littermates (n=9) underwent 48 h of the IVC stenosis. Presented are thrombus weight (i), thrombus length (ii), and thrombosis prevalence (iii). Note absence of thrombosis in both strains of MC-deficient mice and restoration of thrombosis after the adoptive transfer. D, WT (i) or a Kit^{W-sh} (ii) mice underwent IVC stenosis for 48 h, and then, thrombus formation was analyzed by Doppler ultrasound. Note lack of flow in the IVC of the WT mouse and presence of flow in the IVC of the MC-deficient mouse. (iii) Comparison of thrombosis prevalence between WT and MC-deficient mice analyzed by ultrasound.

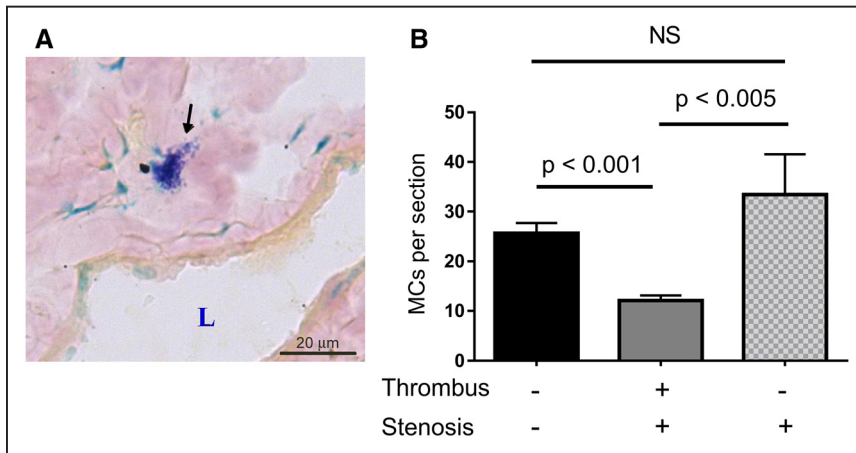


Figure 2. Granule-containing mast cells (MCs) are present in the inferior vena cava (IVC) wall, and their number decreases with thrombosis. **A**, IVCs from wild-type mice were frozen-sectioned and stained with Toluidine blue. MC is indicated by arrow. L, lumen. **B**, MCs were counted in transverse sections of intact IVCs (black bar, n=11), IVCs after 48 h stenosis with a thrombus (gray bar, n=11), and IVCs after 48 h stenosis that did not result in thrombus formation (patterned bar, n=7).

mice administered with compound 48/80 had a thrombus (Figure 3). This result implied that functional MCs are critical for venous thrombosis.

Lack of MCs Is Accompanied by Suppressed Endothelial Activation

We have previously reported the key role of endothelial activation and secretion of WPB constituents for DVT initiation in mice.¹³ Because mice lacking MCs are protected against DVT, we next tested whether MCs regulate activation of the endothelium. Unchallenged Kit^{W-sh} mice had lower plasma levels of vWF (Figure 4A), suggesting an association between presence or absence of MCs and circulating levels of vWF under quiescent conditions. Stenosis of the IVC promotes cell recruitment, which is a prerequisite to thrombus development.^{13,17} Therefore, we next tested whether this process is modified in the absence of MCs. Using intravital microscopy, we demonstrated that in WT mice 6 hours after stenosis application, multiple platelets stay adherent to the IVC wall (Figure 4B; Online Movie I). In contrast, in MC-null mice, only single platelets were adhered with majority of the cells moving with blood flow without detectable contact with the endothelium (Figure 4B; Online Movie II). The area covered by adhered platelets was $6.9 \pm 0.73\%$ in WT mice versus $2.4 \pm 0.54\%$ in Kit^{W-sh} animals ($P < 0.003$; Figure 4C). This result suggests that MCs are implicated in endothelial activation preceding DVT. MC releasate was also able to stimulate

expression of ICAM-1 (intercellular adhesion molecule-1) on HUVECs (human umbilical vein endothelial cells) in vitro (Online Figure IV), which is involved in leukocyte accumulation near the vessel wall. In addition, stenosis induced elevation in plasma levels of sP-selectin, a biomarker of venous thrombosis both in experimental animals and patients,^{17,39-42} in control but not in Kit^{W-sh} mice (Online Figure V). Thus, MC deficiency results in suppressed cell recruitment to the vascular wall at early stages of DVT, potentially because of impaired release of adhesive substances from the endothelium.

Prevention of MC Degranulation Is Protective Against DVT

MCs fulfill their biological functions largely through release of their granule components. Drugs stabilizing MC membrane preventing degranulation are on the market and used predominantly to treat allergic diseases. We next studied whether this pharmacological approach could be beneficial for DVT prophylaxis. Systemic administration of sodium cromoglycate (cromolyn) or ketotifen, MC membrane stabilizers, exerted an antithrombotic effect in the DVT model (Figure 5). Thrombosis prevalence fell from 71% in saline-treated controls to 31% in animals that received cromolyn ($P < 0.04$) and from 80% to 14% in mice that were administered ketotifen ($P < 0.02$). Thus, MC membrane stabilization can potentially be considered as an anti-DVT approach.

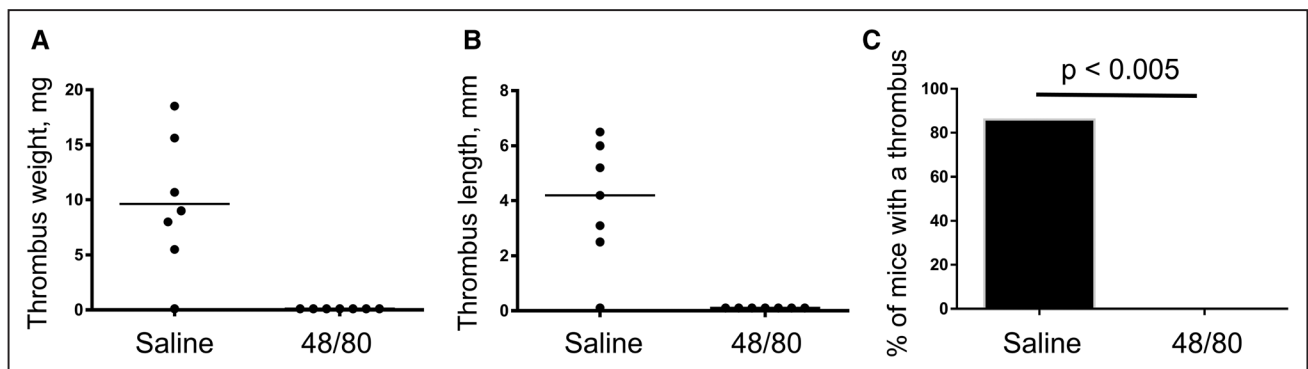


Figure 3. Mast cell (MC) granule depletion prevents deep vein thrombosis. MC granules were depleted by consecutive daily injections of the compound 48/80 as described in the Methods section. Control mice were administered sterile saline. Then, mice were subjected to 48 h inferior vena cava stenosis, and thrombi were excised and measured. n=7 in both groups. Presented are thrombus weight (A), thrombus length (B), and thrombosis prevalence (C).

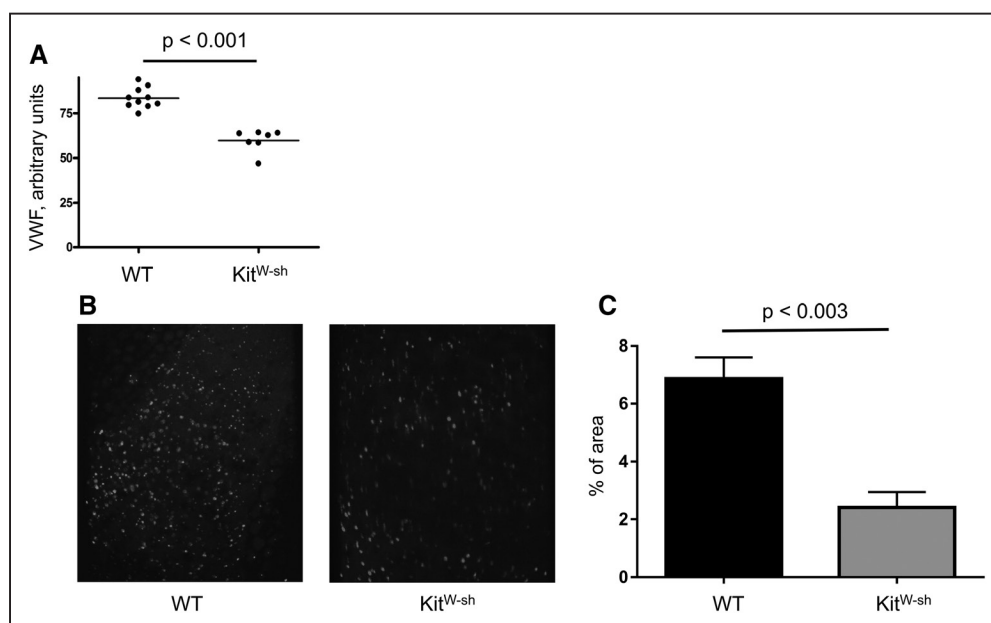


Figure 4. Mast cells regulate plasma levels of von Willebrand Factor (vWF) and platelet recruitment to the stenosed inferior vena cava (IVC) wall. **A**, Blood was drawn from the retro-orbital plexus of unchallenged wild-type (WT) and $\text{Kit}^{\text{W-sh}}$ mice ($n=7-10$), stabilized with sodium citrate, and plasma levels of vWF were measured. **B**, Representative composite images of 10 consecutive frames of intravital microscopy showing fluorescent platelets adhered to the IVC wall 6 h after stenosis induction. Only mice without a thrombus at this time point were used. **C**, Percent of the area covered by adhered platelets in WT (black bar) and $\text{Kit}^{\text{W-sh}}$ mice 6 h after stenosis application. $n=4$ for both groups.

Topical application of compound 48/80 or histamine potentiates thrombosis in WT and induces DVT in $\text{Kit}^{\text{W-sh}}$ mice.

MCs exacerbate DVT apparently by inducing endothelial activation and release of WPB. To further explore mechanisms, through which MCs exert their prothrombotic effect,

we performed topical application of MC secretagogue compound 48/80 or histamine, a potent secretagogue of WPB uniquely synthesized and stored in MCs. Six hours of IVC stenosis resulted in thrombosis in 2 out of 8 (25%) WT control mice (Figure 6). In contrast, WT mice locally treated

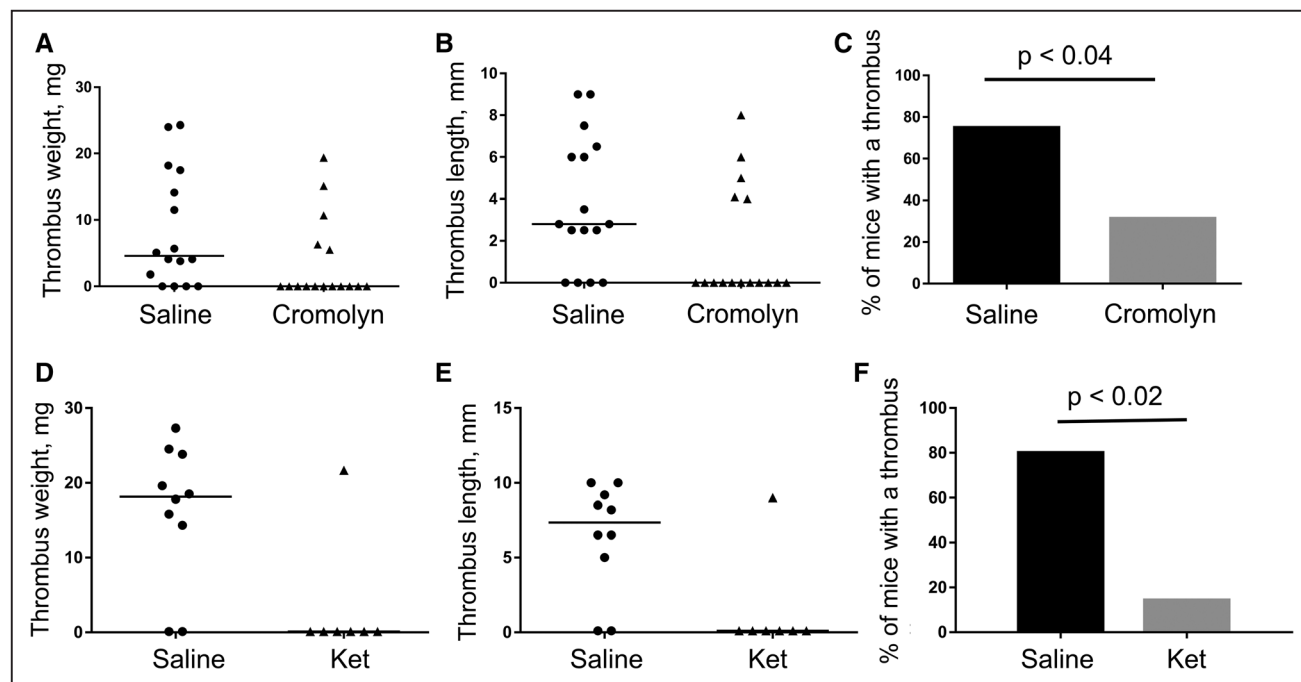


Figure 5. Inhibition of mast cell degranulation by membrane stabilization protects against deep vein thrombosis (DVT). **A–C**, Wild-type (WT) mice were administered sodium cromoglycate IP (cromolyn, 100 mg/kg body weight, $n=17$) or sterile saline ($n=16$) 24 h and 30 minutes before inferior vena cava stenosis induction. **D–F**, WT mice were injected with ketotifen IP (25 mg/kg body weight, $n=7$) or sterile saline ($n=10$) 24 h and 30 minutes before and 24 h after DVT surgery. Thrombus formation was checked 48 h later. Thrombus weight (**A** and **D**), thrombus length (**B** and **E**), and thrombosis prevalence (**C** and **F**) are shown.

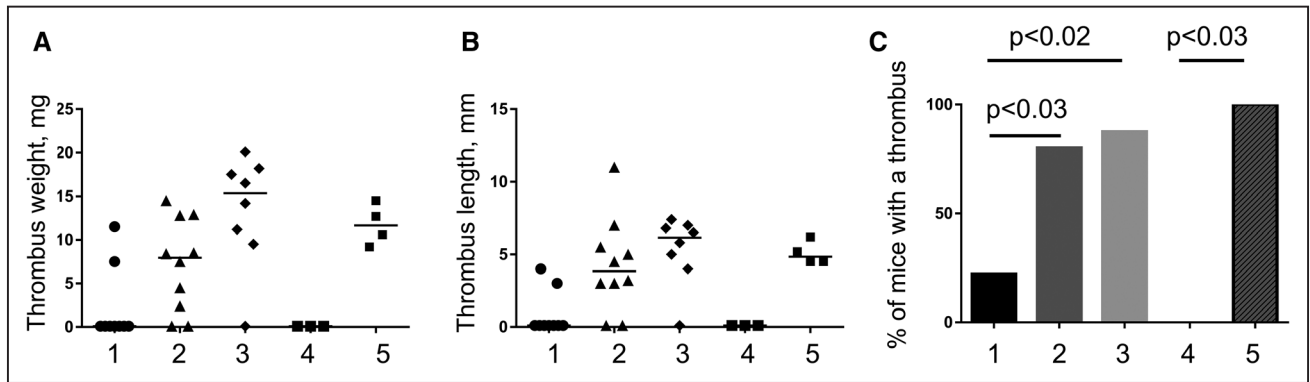


Figure 6. Topical application of compound 48/80 or histamine potentiates thrombosis in wild type (WT) and histamine induces deep vein thrombosis in Kit^{W-sh} mice. WT and Kit^{W-sh} mice underwent inferior vena cava (IVC) stenosis. During surgery, a strip of paper soaked in saline or solutions of either compound 48/80 (in WT mice only) or histamine (in both WT and Kit^{W-sh} mice) was applied to the IVC for 10 min immediately after stenosis induction. In 6 h, mice were euthanized, and thrombus formation tested. The groups are: 1, WT+saline; 2, WT+48/80; 3, WT+histamine; 4, Kit^{W-sh}+saline; and 5, Kit^{W-sh}+histamine. Presented are thrombus weight (A), thrombus length (B), and thrombosis prevalence (C).

with compound 48/80 developed thrombus in 80% of cases ($P < 0.03$). Histamine-treated WT mice developed a thrombus in 87.5% of cases ($P < 0.02$ versus saline-treated animals). Interestingly, all Kit^{W-sh} animals, normally completely protected against DVT even at longer time points (48 hours), had thrombi after topical application of histamine, indicating that histamine can correct DVT defect in MC-deficient mice. Thus, it seems that local activation of MCs in WT mice promotes DVT, and this effect is likely mediated by histamine release.

We next performed administration of histamine H1 receptor (which is expressed on endothelial cells^{43,44}) antagonists, pyrilamine maleate or cetirizine, to test whether inhibition of histamine effect can inhibit thrombosis. No protective effect of either of the substances on DVT was observed (Online Figure VI).

Discussion

Current methods of DVT prophylaxis, targeting clotting factors, are inevitably accompanied by bleeding complications in a certain percent of patients. This is not surprising because components of blood coagulation system are involved in normal hemostasis, and downregulation of their function predictably affects physiological bleeding arrest. We report here (1) a novel role of MCs in DVT and (2) an attempt to prevent venous thrombosis using an alternative approach—inhibition of MCs, which are not directly implicated in hemostasis and therefore targeting them is unlikely to cause pathological bleeding.

Venous thrombus formation induced by flow distortion is based on endothelial activation, release of WPB components, and recruitment of innate immune cells and platelets, which precede thrombosis.^{13,17} Prevention of recruitment of immune cell, for example, in P-selectin-deficient mice, or platelets, results in strong protection against DVT.^{13,17} Thus, mechanisms of DVT in this model include not only blood coagulation but also elements of sterile inflammation. MCs contain both inhibitors of coagulation, such as heparin and tPA, which could be expected to inhibit thrombosis, and proinflammatory mediators, such as histamine and tumor necrosis factor- α , which could exert a prothrombotic effect given that DVT is an immunothrombosis and its mechanisms involve inflammatory components.⁴⁵ This made it initially difficult to predict which

of the pathways would turn out to be more important for DVT. We demonstrate that 2 independent strains of MC-deficient mice were completely protected against DVT in the IVC stenosis model. Lack of thrombosis in the absence of MCs suggests that MCs contain an entity, whose prothrombotic potential outweighs the antithrombotic power of one or several other MC components. One potential explanation of the relative inactivity of MC-derived anticoagulants is that MCs are located in the tissue surrounding the IVC, and their released granule constituents may reach abluminal part of endothelium but not the blood stream. In this scenario, proinflammatory mediators can activate endothelium, whereas heparin and tPA may not exert their anticoagulant effect. It should be noted that a small proportion of WT mice may not produce thrombi in this model. This suggests that the prothrombotic activity of MCs is not absolute and because of unclear reason might not suffice for thrombus formation in certain cases. Also, it should be noted that our results have been obtained on a mouse model, which has substantial differences from the human situation. In particular, murine IVC does not contain valves, whereas in humans, DVT develops inside venous valves. Also, humans have vertical spinal orientation, whereas mice have horizontal one, which means importance of muscle pump in venous return of blood to the heart in humans and unimportance of it in mice. There are difference also in various aspects of the immune system.⁴⁶ For example, neutrophils constitute different percent of leukocytes in mice in humans,^{46,47} which might be relevant to venous thrombosis as neutrophils support DVT by expelling neutrophil extracellular traps.¹⁶ Thus, the validity of our results for humans remains to be verified in future translational studies in patients. The availability of MC membrane stabilizers on the market makes such studies possible in the not too distant future.

Mice used in this study lack MCs because of different mutations in c-Kit (CD117), a receptor to stem cell factor. In addition to absence of MCs, mutations in c-Kit may lead to other abnormalities. For example, homozygous Kit^{W-v} mice have macrocytic anemia and germ cell developmental defects, whereas Kit^{W-sh} mutation is accompanied by abnormal vascular permeability induced by bee venom phospholipase A2. Other

still unknown MC-unrelated defects also cannot be excluded. To rule out alternative explanations of the phenotype in these mice, other than lack of MCs, we performed adoptive transfer of in vitro differentiated MCs into MC-deficient Kit^{W-sh} mice. This procedure has been shown to restore pool of MCs in different organs within 2 to 3 months.^{37,38} Adoptive transfer of MCs restored thrombosis in Kit^{W-sh} mice. Thus, the antithrombotic phenotype results primarily from absence of MCs and not from other defect caused by c-Kit mutation. Although engraftment of transferred MCs might differ from normal MC distribution and pattern of MC pool restoration in the IVC wall is unclear, such abnormal engraftment could be a potential explanation in case of lack of the phenotype recovery rather than in case of successful restoration of thrombosis by MC infusion.

Connective tissue MCs are usually located near blood vessels.⁴⁸ The IVC turned out to be no exception as MCs were found in its wall in the vicinity to abluminal part of the endothelium. Induction of stenosis, which eventually resulted in thrombus formation, was associated with decreased numbers of granule-containing (toluidine blue-positive) MCs per section, whereas MC quantities remained unchanged after stenosis that because of unknown reasons did not lead to thrombosis. This result suggests a link between MC function and DVT development. It is unclear though whether MCs migrate out of the IVC wall or fully degranulate becoming invisible in Toluidine blue staining. Disappearance of MCs specifically in thrombosed IVCs implies that their released cargo triggers DVT.

In addition to genetic mutations, MCs in mice can be targeted pharmacologically. Chronic administration of MC secretagogue, compound 48/80, makes MCs temporarily functionally inactive because of exhaustion of intracellular pools of mediators.³¹ A similar effect can be achieved by MC membrane stabilizers that prevent the liberation of MC granule content.³² We tested both approaches, and they protected mice from DVT. In case of compound 48/80, the antithrombotic effect might also be attributed to downregulation of coagulation factor VII activation,⁴⁹ and, therefore, in addition to its effect on MCs, the compound may suppress also the coagulation cascade. Thrombin generation is important for DVT in this model because low-molecular-weight heparin prevents thrombosis.¹⁷ Partial but significant protection by inhibiting MC degranulation is of particular importance because multiple drugs of this kind are already on the market and used for antiallergic purposes. Sodium cromoglycate used as a MC stabilizer does not have an antiplatelet effect, as shown by platelet function analyzer-100,⁵⁰ and, therefore, platelet inhibition as a mechanism of DVT prevention can be ruled out.

Thus, MCs maintain vWF secretion in unchallenged mice (Figure 4A) and exacerbate DVT likely by stimulating activation of the endothelium and promoting WPB release. Failure to recruit platelets in the intravital microscopy in mice lacking MCs supports this statement. Platelet recruitment is a prerequisite for DVT and is dependent on vWF release from WPBs.^{13,17} The phenotype of MC-deficient animals is similar to vWF knockout mice: in both Kit^{W-sh} and vWF knockout mice, platelet accrual at the site of future thrombus is decreased. Besides platelets, leukocytes are also recruited to the IVC wall after stenosis and facilitate DVT by releasing neutrophil extracellular traps.¹⁶ Also, MC releasate was able to stimulate expression of ICAM-1, a major adhesion receptor

for leukocytes, on endothelial cells in vitro. Consequently, MCs upregulate endothelial activation and liberation of vWF and P-selectin from WPBs and by this support DVT.

Interestingly, IVC stenosis is accompanied by elevated plasma sP-selectin levels, which can originate from both platelets and endothelium. Regardless of sP-selectin origin in this particular model, its plasma content is also regulated by MCs because in Kit^{W-sh} mice no sP-selectin upregulation was observed. Plasma sP-selectin is known to promote DVT in mice and is a DVT biomarker^{39,42,51} so that its increase in WT mice after IVC stenosis further suggests the similarity between the animal model and the human disease.

Topical application of MC secretagogue compound 48/80 stimulated thrombosis, suggesting that it is a MC granule constituent, likely a small molecule that affects endothelium or can permeate into blood, which promotes DVT in this model. Histamine is the likely candidate for stimulating WPB release. It is a potent WPB secretagogue,⁵² and MCs are a unique source of it. Local application of histamine to the IVC accelerated thrombosis in WT mice and induced DVT in MC-deficient mice. Thus, histamine is able to overcome the effect of MC deficiency, suggesting that it is the important constituent of MC granules, which may mediate MC prothrombotic effect. However, systemic administration of histamine H1 receptor inhibitors did not protect mice from DVT. This may be explained in several ways. First, histamine may not be a decisive WPB component, and the prothrombotic effect of MCs may require combined action of ≥ 2 additional components (eg, tumor necrosis factor- α). Second, histamine may act through >1 receptor. For example, H2 receptor operates in some vascular beds⁵³ so inhibition of only 1 receptor might be insufficient. Third, histamine from MC granules reaches endothelial cells from their basal side, and, therefore, its effect may not be fully abolished by the receptor inhibitors, which operate at the luminal side. Given relatively long time of the experiment (48 hours), insufficient circulation time of the inhibitors also cannot be ruled out.

If MC absence or suppression protects against thrombosis, a question may arise as to why mastocytosis does not lead to thrombus formation. For example, no thrombi in the skin vessels of urticaria pigmentosa patients have been described. The explanation of this phenomenon may be that when the number of MCs increases, the ratio between anticoagulants (heparin, tPA) and proinflammatory mediators (histamine, tumor necrosis factor- α) released from MCs shifts toward the anticoagulants, rendering the resulting phenotype rather normal or bleeding than prothrombotic.

In conclusion, we have demonstrated that MCs are potent regulators of DVT in mice. The effect of MCs is likely to be at least partially implemented through release of histamine, although other MC granule constituents may contribute as well. Kit^{W-sh} mice have normal bleeding time, implying that temporary switching off MCs should not result in excessive bleeding. Thus, MCs may be considered a promising target for DVT prevention in humans.

Acknowledgments

We thank Nashitha Kabir for technical assistance with ultrasound Doppler. We also thank Dr P. Harrison and Dr C. Gardiner for their help with murine coagulation tests.

Sources of Funding

The work was supported by the British Heart Foundation (PG/13/60/30406) and the University of Birmingham to A. Brill and by the National Institutes of Health (5R01HL102101) to D.D. Wagner.

Disclosures

None.

References

- Heit JA, Spencer FA, White RH. The epidemiology of venous thromboembolism. *J Thromb Thrombolysis*. 2016;41:3–14. doi: 10.1007/s11239-015-1311-6.
- Silverstein MD, Heit JA, Mohr DN, Petterson TM, O'Fallon WM, Melton LJ III. Trends in the incidence of deep vein thrombosis and pulmonary embolism: a 25-year population-based study. *Arch Intern Med*. 1998;158:585–593.
- Becattini C, Agnelli G, Lankeit M, Masotti L, Pruszczyk P, Casazza F, Vanni S, Nitti C, Kamphuisen P, Vedovati MC, De Natale MG, Konstantinides S. Acute pulmonary embolism: mortality prediction by the 2014 European society of cardiology risk stratification model. *Eur Respir J*. 2016;48:780–786. doi: 10.1183/13993003.00024-2016.
- Office of the Surgeon General (US). *The Surgeon General's Call to Action to Prevent Deep Vein Thrombosis and Pulmonary Embolism*. National Heart, Lung, and Blood Institute. Rockville, MD; 2008.
- Thom T, Haase N, Rosamond W, et al; American Heart Association Statistics Committee and Stroke Statistics Subcommittee. Heart disease and stroke statistics—2006 update: a report from the American Heart Association Statistics Committee and Stroke Statistics Subcommittee. *Circulation*. 2006;113:e85–e151. doi: 10.1161/CIRCULATIONAHA.105.171600.
- Ashrani AA, Heit JA. Incidence and cost burden of post-thrombotic syndrome. *J Thromb Thrombolysis*. 2009;28:465–476. doi: 10.1007/s11239-009-0309-3.
- Condliffe R, Kiely DG, Gibbs JS, Corris PA, Peacock AJ, Jenkins DP, Goldsmith K, Coghlan JG, Pepke-Zaba J. Prognostic and aetiological factors in chronic thromboembolic pulmonary hypertension. *Eur Respir J*. 2009;33:332–338. doi: 10.1183/09031936.00092008.
- Goldhaber SZ. Risk factors for venous thromboembolism. *J Am Coll Cardiol*. 2010;56:1–7. doi: 10.1016/j.jacc.2010.01.057.
- Enga KF, Rye-Holmboe I, Hald EM, Løchen ML, Mathiesen EB, Njølstad I, Wilsgaard T, Braekkan SK, Hansen JB. Atrial fibrillation and future risk of venous thromboembolism: the Tromsø study. *J Thromb Haemost*. 2015;13:10–16. doi: 10.1111/jth.12762.
- Sevitt S. The structure and growth of valve-pocket thrombi in femoral veins. *J Clin Pathol*. 1974;27:517–528.
- Bovill EG, van der Vliet A. Venous valvular stasis-associated hypoxia and thrombosis: what is the link? *Annu Rev Physiol*. 2011;73:527–545. doi: 10.1146/annurev-physiol-012110-142305.
- Hamer JD, Malone PC, Silver IA. The PO2 in venous valve pockets: its possible bearing on thrombogenesis. *Br J Surg*. 1981;68:166–170.
- Brill A, Fuchs TA, Chauhan AK, Yang JJ, De Meyer SF, Köllnberger M, Wakefield TW, Lämmle B, Massberg S, Wagner DD. von Willebrand factor-mediated platelet adhesion is critical for deep vein thrombosis in mouse models. *Blood*. 2011;117:1400–1407. doi: 10.1182/blood-2010-05-287623.
- Myers DD, Wakefield TW. Inflammation-dependent thrombosis. *Front Biosci*. 2005;10:2750–2757.
- Reitsma PH, Versteeg HH, Middeldorp S. Mechanistic view of risk factors for venous thromboembolism. *Arterioscler Thromb Vasc Biol*. 2012;32:563–568. doi: 10.1161/ATVBAHA.111.242818.
- Brill A, Fuchs TA, Savchenko AS, Thomas GM, Martinod K, De Meyer SF, Bhandari AA, Wagner DD. Neutrophil extracellular traps promote deep vein thrombosis in mice. *J Thromb Haemost*. 2012;10:136–144. doi: 10.1111/j.1538-7836.2011.04544.x.
- von Brühl ML, Stark K, Steinhart A, et al. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. *J Exp Med*. 2012;209:819–835. doi: 10.1084/jem.20112322.
- Gurish MF, Austen KF. Developmental origin and functional specialization of mast cell subsets. *Immunity*. 2012;37:25–33. doi: 10.1016/j.immuni.2012.07.003.
- Lippi G, Favalaro EJ. Allergy and venous thromboembolism: a casual or causative association. *Semin Thromb Hemost*. 2016;42:63–68. doi: 10.1055/s-0035-1568876.
- Modena BD, Dazy K, White AA. Emerging concepts: mast cell involvement in allergic diseases. *Transl Res*. 2016;174:98–121. doi: 10.1016/j.trsl.2016.02.011.
- Voehringer D. Protective and pathological roles of mast cells and basophils. *Nat Rev Immunol*. 2013;13:362–375. doi: 10.1038/nri3427.
- Sillaber C, Baghestanian M, Bevec D, Willheim M, Agis H, Kapiotis S, Füreder W, Bankl HC, Kiener HP, Speiser W, Binder BR, Lechner K, Valent P. The mast cell as site of tissue-type plasminogen activator expression and fibrinolysis. *J Immunol*. 1999;162:1032–1041.
- Kunder CA, St John AL, Abraham SN. Mast cell modulation of the vascular and lymphatic endothelium. *Blood*. 2011;118:5383–5393. doi: 10.1182/blood-2011-07-358432.
- Bankl HC, Grossschmidt K, Pikula B, Bankl H, Lechner K, Valent P. Mast cells are augmented in deep vein thrombosis and express a profibrinolytic phenotype. *Hum Pathol*. 1999;30:188–194.
- Nocka K, Tan JC, Chiu E, Chu TY, Ray P, Traktman P, Besmer P. Molecular bases of dominant negative and loss of function mutations at the murine c-kit/white spotting locus: W37, Wv, W41 and W. *EMBO J*. 1990;9:1805–1813.
- Lyon MF, Glenister PH. A new allele sash (Wsh) at the W-locus and a spontaneous recessive lethal in mice. *Genet Res*. 1982;39:315–322.
- Stevens J, Loutit JF. Mast cells in spotted mutant mice (W Ph mi). *Proc R Soc Lond B Biol Sci*. 1982;215:405–409.
- Geddings J, Aleman MM, Wolberg A, von Brühl ML, Massberg S, Mackman N. Strengths and weaknesses of a new mouse model of thrombosis induced by inferior vena cava stenosis: communication from the SSC of the ISTH. *J Thromb Haemost*. 2014;12:571–573. doi: 10.1111/jth.12510.
- Shelburne CP, McLachlan JB, Abraham SN. In vivo models for studying mast cell-dependent responses to bacterial infection. *Methods Mol Biol*. 2006;315:363–381.
- Kuehn HS, Rädinger M, Gilfillan AM. Measuring mast cell mediator release. *Curr Protoc Immunol*. 2010;Chapter 7:Unit7.38. doi: 10.1002/0471142735.im0738s91.
- Carvalho M, Benjamim C, Santos F, Ferreira S, Cunha F. Effect of mast cells depletion on the failure of neutrophil migration during sepsis. *Eur J Pharmacol*. 2005;525:161–169. doi: 10.1016/j.ejphar.2005.09.049.
- Ramos L, Peña G, Cai B, Deitch EA, Ulloa L. Mast cell stabilization improves survival by preventing apoptosis in sepsis. *J Immunol*. 2010;185:709–716. doi: 10.4049/jimmunol.1000273.
- Bot I, de Jager SC, Zernecke A, Lindstedt KA, van Berkel TJ, Weber C, Biessen EA. Perivascular mast cells promote atherogenesis and induce plaque destabilization in apolipoprotein E-deficient mice. *Circulation*. 2007;115:2516–2525. doi: 10.1161/CIRCULATIONAHA.106.660472.
- Fabritz L, Hoogendijk MG, Scicluna BP, et al. Load-reducing therapy prevents development of arrhythmogenic right ventricular cardiomyopathy in plakoglobin-deficient mice. *J Am Coll Cardiol*. 2011;57:740–750. doi: 10.1016/j.jacc.2010.09.046.
- Sabrane K, Kruse MN, Fabritz L, Zetsche B, Mitko D, Skryabin BV, Zwienen M, Baba HA, Yanagisawa M, Kuhn M. Vascular endothelium is critically involved in the hypotensive and hypovolemic actions of atrial natriuretic peptide. *J Clin Invest*. 2005;115:1666–1674. doi: 10.1172/JCI23360.
- Ni H, Denis CV, Subbarao S, Degen JL, Sato TN, Hynes RO, Wagner DD. Persistence of platelet thrombus formation in arterioles of mice lacking both von Willebrand factor and fibrinogen. *J Clin Invest*. 2000;106:385–392. doi: 10.1172/JCI9896.
- Wolters PJ, Mallen-St Clair J, Lewis CC, Villalta SA, Baluk P, Erle DJ, Caughey GH. Tissue-selective mast cell reconstitution and differential lung gene expression in mast cell-deficient Kit(W-sh)/Kit(W-sh) sash mice. *Clin Exp Allergy*. 2005;35:82–88. doi: 10.1111/j.1365-2222.2005.02136.x.
- Furuta T, Kikuchi T, Iwakura Y, Watanabe N. Protective roles of mast cells and mast cell-derived TNF in murine malaria. *J Immunol*. 2006;177:3294–3302.
- Antonopoulos CN, Sfyroeras GS, Kakisis JD, Moulakakis KG, Liapis CD. The role of soluble P selectin in the diagnosis of venous thromboembolism. *Thromb Res*. 2014;133:17–24. doi: 10.1016/j.thromres.2013.08.014.
- Vandy FC, Stabler C, Eliassen AM, Hawley AE, Guire KE, Myers DD, Henke PK, Wakefield TW. Soluble P-selectin for the diagnosis of lower extremity deep venous thrombosis. *J Vasc Surg Venous Lymphat Disord*. 2013;1:117–125. doi: 10.1016/j.jvsv.2012.09.001.
- Culmer DL, Diaz JA, Hawley AE, Jackson TO, Shuster KA, Sigler RE, Wakefield TW, Myers DD, Jr. Circulating and vein wall P-selectin promote venous thrombogenesis during aging in a rodent model. *Thromb Res*. 2013;131:42–48. doi: 10.1016/j.thromres.2012.10.013.

42. Ramacciotti E, Blackburn S, Hawley AE, Vandy F, Ballard-Lipka N, Stabler C, Baker N, Guire KE, Rectenwald JE, Henke PK, Myers DD, Jr, Wakefield TW. Evaluation of soluble P-selectin as a marker for the diagnosis of deep venous thrombosis. *Clin Appl Thromb Hemost*. 2011;17:425–431. doi: 10.1177/1076029611405032.
43. Hill SJ, Ganellin CR, Timmerman H, Schwartz JC, Shankley NP, Young JM, Schunack W, Levi R, Haas HL. International Union of Pharmacology. XIII. Classification of histamine receptors. *Pharmacol Rev*. 1997;49:253–278.
44. Li H, Burkhardt C, Heinrich UR, Brausch I, Xia N, Förstermann U. Histamine upregulates gene expression of endothelial nitric oxide synthase in human vascular endothelial cells. *Circulation*. 2003;107:2348–2354. doi: 10.1161/01.CIR.0000066697.19571.AF.
45. Engelmann B, Massberg S. Thrombosis as an intravascular effector of innate immunity. *Nat Rev Immunol*. 2013;13:34–45. doi: 10.1038/nri3345.
46. Mestas J, Hughes CC. Of mice and not men: differences between mouse and human immunology. *J Immunol*. 2004;172:2731–2738.
47. Doeing DC, Borowicz JL, Crockett ET. Gender dimorphism in differential peripheral blood leukocyte counts in mice using cardiac, tail, foot, and saphenous vein puncture methods. *BMC Clin Pathol*. 2003;3:3. doi: 10.1186/1472-6890-3-3.
48. Metcalfe DD, Baram D, Mekori YA. Mast cells. *Physiol Rev*. 1997;77:1033–1079.
49. Chu AJ, Wang ZG, Fox MJ. III. Instantaneous inhibition by compound 48/80 of tissue factor-initiated extrinsic coagulation is mediated by the downregulation of factor VII activation. *Arch Biochem Biophys*. 2000;377:357–365. doi: 10.1006/abbi.2000.1771.
50. Nemmar A, Hoet PH, Vermeylen J, Nemery B, Hoylaerts MF. Pharmacological stabilization of mast cells abrogates late thrombotic events induced by diesel exhaust particles in hamsters. *Circulation*. 2004;110:1670–1677. doi: 10.1161/01.CIR.0000142053.13921.21.
51. Myers DD, Jr, Rectenwald JE, Bedard PW, Kaila N, Shaw GD, Schaub RG, Farris DM, Hawley AE, Wroblewski SK, Henke PK, Wakefield TW. Decreased venous thrombosis with an oral inhibitor of P selectin. *J Vasc Surg*. 2005;42:329–336. doi: 10.1016/j.jvs.2005.04.045.
52. Hamilton KK, Sims PJ. Changes in cytosolic Ca²⁺ associated with von Willebrand factor release in human endothelial cells exposed to histamine. study of microcarrier cell monolayers using the fluorescent probe indo-1. *J Clin Invest*. 1987;79:600–608. doi: 10.1172/JCI112853.
53. Hekimian G, Côte S, Van Sande J, Boeynaems JM. H2 receptor-mediated responses of aortic endothelial cells to histamine. *Am J Physiol*. 1992;262:H220–H224.

Circulation Research

JOURNAL OF THE AMERICAN HEART ASSOCIATION



Mast Cells Granular Contents Are Crucial for Deep Vein Thrombosis in Mice Tatyana Ponomaryov, Holly Payne, Larissa Fabritz, Denisa D. Wagner and Alexander Brill

Circ Res. 2017;121:941-950; originally published online July 24, 2017;
doi: 10.1161/CIRCRESAHA.117.311185

Circulation Research is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2017 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7330. Online ISSN: 1524-4571

The online version of this article, along with updated information and services, is located on the
World Wide Web at:

<http://circres.ahajournals.org/content/121/8/941>

Free via Open Access

Data Supplement (unedited) at:

<http://circres.ahajournals.org/content/suppl/2017/07/24/CIRCRESAHA.117.311185.DC1>

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Circulation Research* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the [Permissions and Rights Question and Answer](#) document.

Reprints: Information about reprints can be found online at:
<http://www.lww.com/reprints>

Subscriptions: Information about subscribing to *Circulation Research* is online at:
<http://circres.ahajournals.org/subscriptions/>

SUPPLEMENTAL MATERIAL

Detailed Methods

Adoptive transfer of MCs into Kit^{W-sh} mice. Ten millions of *in vitro* differentiated MCs were injected into 3-4 weeks old Kit^{W-sh} mice through the tail vein. In 8 – 10 weeks, mice were subjected to 48 h IVC stenosis.

Toluidine blue staining and microscopy for mast cells granule content. Frozen sections were submerged in absolute ethanol, washed with running water and subjected to staining in Toluidine blue (TB) solution for 1 min. The staining solution was prepared freshly from 5 ml stock of TB (1% TB (Sigma) in 70% ethanol) combined with 45 ml fresh 1% NaCl solution in water (pH 2.0-2.5). After staining sections were washed with running water, submerged in 95% and 100% ethanol, soaked in a clearant and mounted with hydrophobic mount. Images were taken by Axio Scan.Z1 (Zeiss).

Local application of compound 48-80 or histamine. After stenosis application, a strip of Whatman paper soaked in solution of compound 48-80 (1 mg/ml), histamine (100 mg/ml; both from Sigma) or saline (control) was placed on the IVC for 10 min. Then peritoneum and skin were closed and mice were euthanized in 6 h to test the development of thrombi. Due to manual fashion of the paper strip application small variation in experimental conditions might take place from mouse to mouse.

Intravital microscopy of the stenosed IVC site and evaluation of the space covered with immobilized platelets. Mice were subjected to IVC stenosis for 6 h. Animals that had thrombi, as judged by observation through the IVC wall and inability to squeeze the IVC with forceps, were excluded from the experiment. Thereafter, mice were anesthetized with tribromethanol and syngeneic washed platelets labeled by calcein AM (from Sigma; 2.5% of the total amount of circulating platelets) were infused intravenously. Then the IVC was exposed and a coverslip was placed on its top. Fluorescent platelets were visualized 1-2 mm below (in caudal direction) the ligation site using 3i VIVO-SDC confocal system with Yokogawa CSU-10 and Photometrics Evolve EMCCD camera on an Olympus BX61WI upright microscope with air objective x10. Focus was adjusted to the upper focal plane. Flow was recorded for at least 1 min. Fiji/ImageJ33 software was used to evaluate areas covered by adhered platelets. Median-based intensity projection from 10 randomly chosen consecutive acquired images (corresponding to 1 s) was employed to generate a single projected image. After automatic set of threshold, total area of adhered platelets was measured by a Fiji incorporated function “Analyze particles”.

Treatment with histamine H1 receptor inhibitors. WT mice were administered pyrilamine maleate or cetirizine (both 30 mg/kg) 24 h and 30 min prior to IVC stenosis application, and 24 h thereafter. In 48 h after surgery, thrombosis was visually verified and thrombi were excised for measurement.

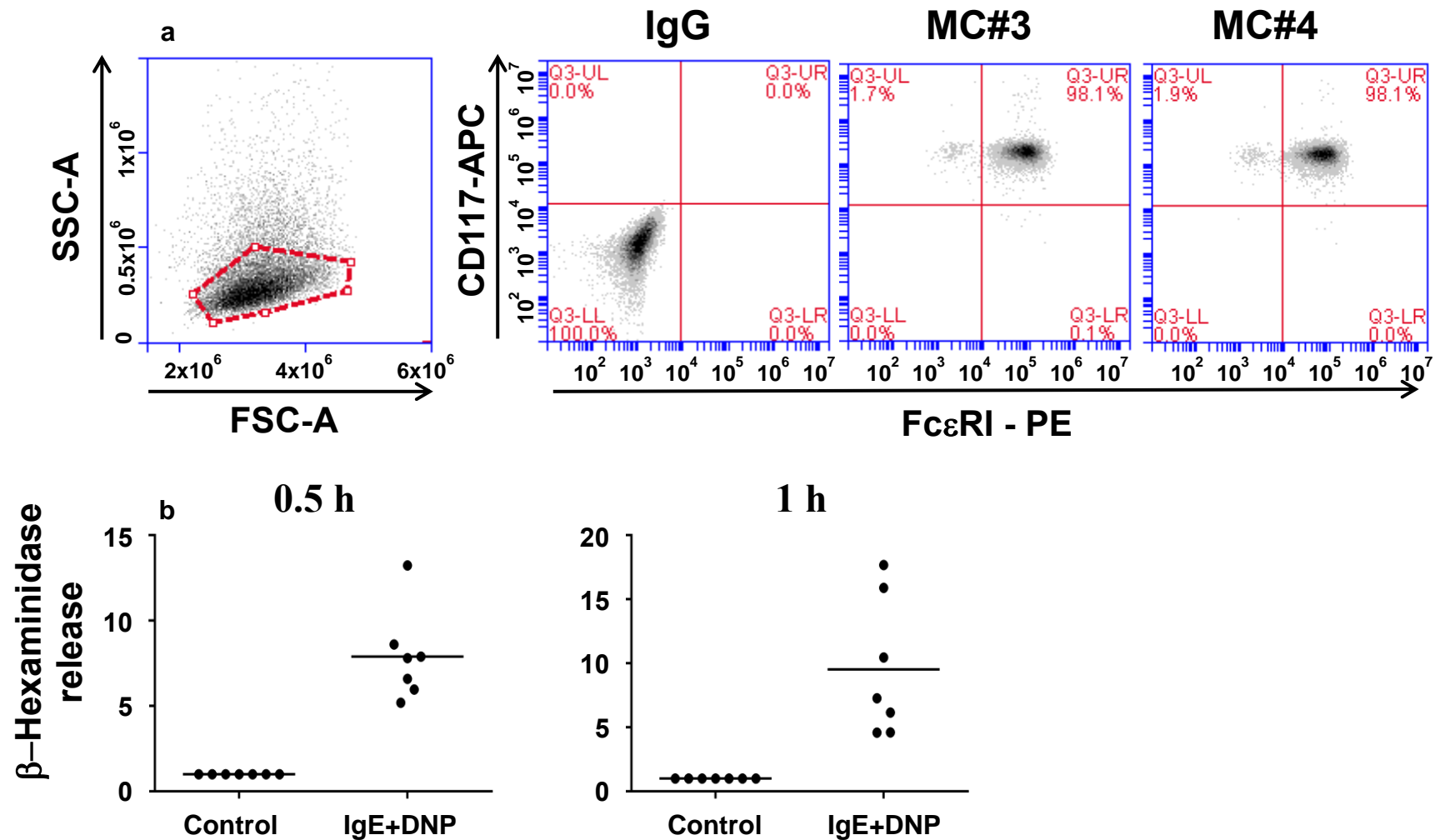
ICAM-1 expression on HUVEC cells. HUVEC cells were purchased from Gibco (Life Technologies Corporation, Carlsbad, CA) and grown according to the supplier instructions. For stimulation experiments, low passage HUVEC cells were plated in 24 wells until they reached confluency. Mast cells were primed overnight with anti-DMB IgE and stimulated with DMB for 6 h. The cleared MC releasate was applied on HUVEC cells for 20 h. After stimulation, cells were collected by Accutase treatment and stained with anti-human ICAM-1 (CD54, clone HA58, eBioscience). Flow cytometry was performed using Accuri C6 cytometer (BD Biosciences, USA).

Measurement of soluble P-selectin (sP-selectin) plasma levels. The content of sP-selectin was determined using ELISA kit (R&D systems) in accordance to manufacturer's instructions.

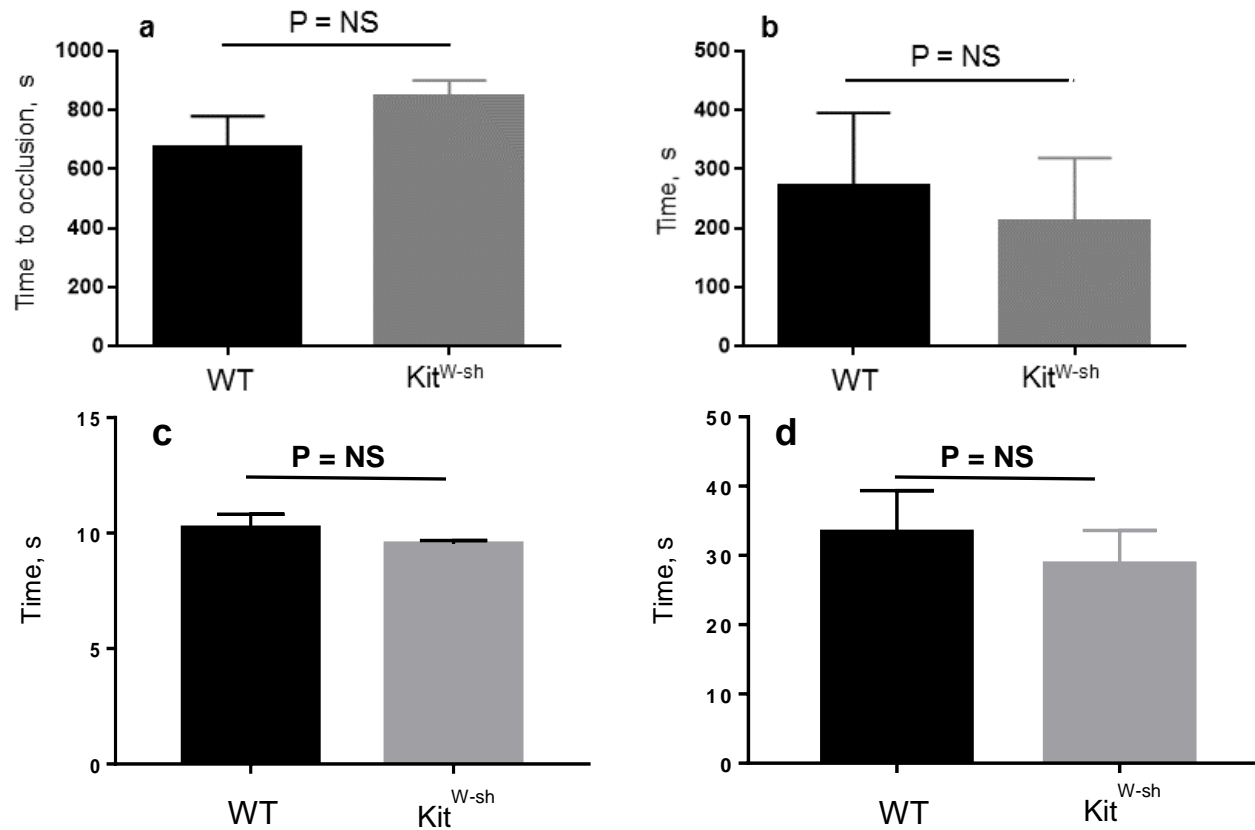
Coagulation tests. Prothrombin time (PT) and activated partial thromboplastin time (aPTT) were performed on Sysmex CS5100 coagulometer. Prothrombin time reagent Thromborel S and aPTT reagent Actin FS were from Siemens.

Statistics. Non-parametric results (e.g., length and weight of thrombi) were compared by the Mann-Whitney test. Parametric data (e.g., area covered by adhered platelets) were compared using two-tailed unpaired Student's t-test or (in case of multiple comparisons) one-way ANOVA. Thrombosis prevalence was compared by contingency table followed by the Fisher's exact square method. Statistics was calculated using GraphPad Prism 7 (GraphPad Software, Inc. La Jolla, CA). Difference between experimental groups was considered significant at $p < 0.05$.

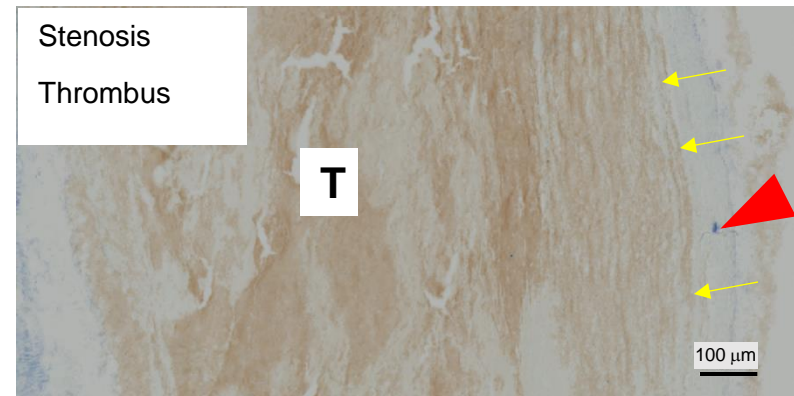
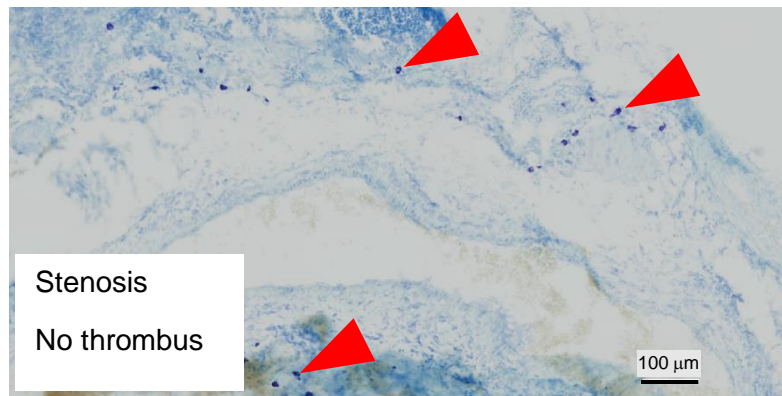
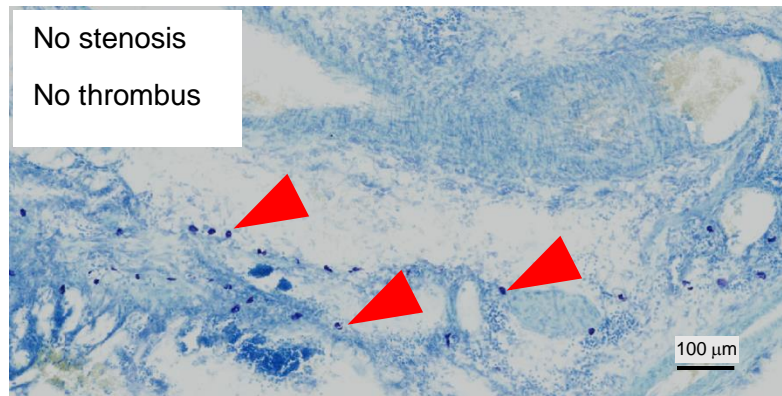
Supplemental Figures



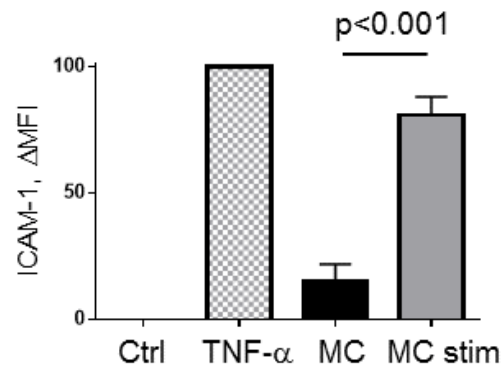
Online Figure I. Evaluation of purity and functionality of the in vitro differentiated MCs. (a) In vitro differentiated MCs were stained for CD117 and FcεRI; more than 90% of the cells expressed both receptors. Two MC batches used in the adoptive transfer experiments are presented (MC#3 and MC#4). **(b)** MC functionality was checked by their ability to secrete granular content upon stimulation. Each dot represents an individual cell line.



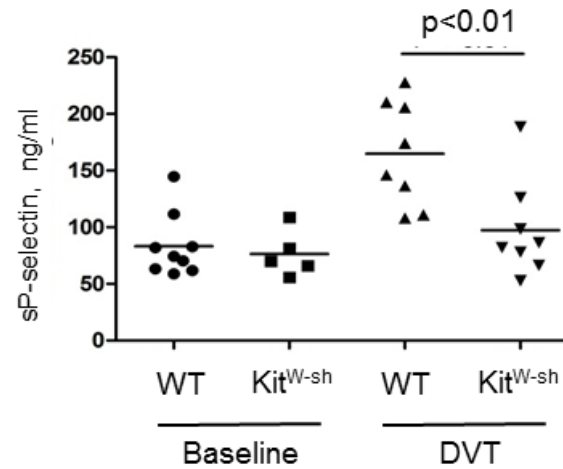
Online Figure II. No difference in normal hemostasis and arterial thrombosis between WT and Kit^{W-sh} mice. (a) Fluorescent congenic platelets were infused into anesthetized WT or Kit^{W-sh} mice (n=4 to 5 for each group), their mesentery was exposed and a strip of paper soaked in 10% ferric chloride was applied to the arteriole for 5 min. Time to full vessel occlusion was recorded (b) Tail tip (3 mm) was cut in anesthetized WT or Kit^{W-sh} mice (n=6 to 8 for each group), placed into a test tube with warm saline and time to complete bleeding arrest was recorded. PT (c) and aPTT (d) tests were performed using a coagulometer (n = 4 to 5 mice per group).



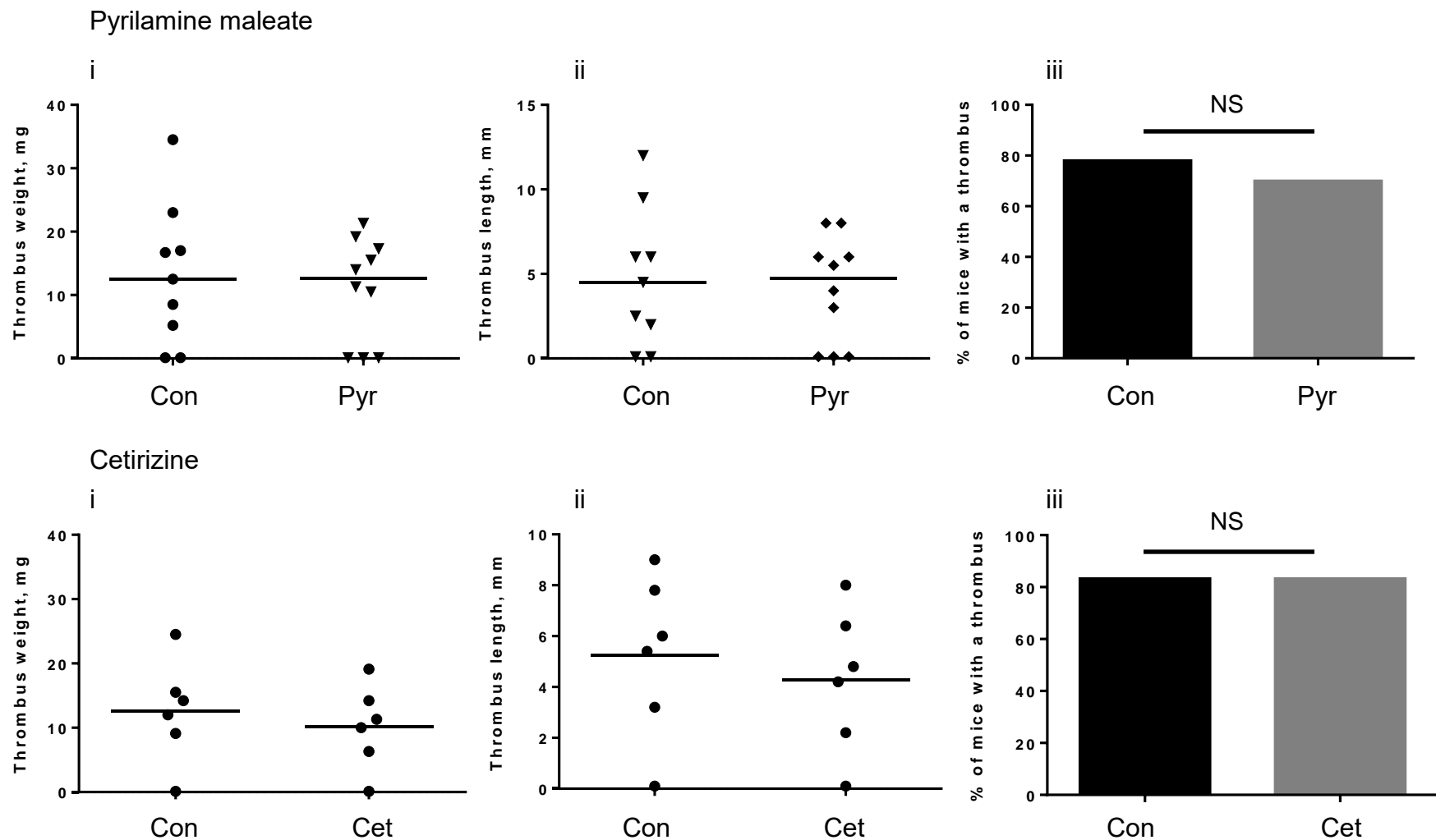
Online Figure III. Representative images of MCs in the vicinity of the IVC. IVC was excised from unchallenged mice (upper left) or mice after 48 h stenosis with (upper right) or without (upper right) a thrombus. Mast cells were stained with toluidine blue. Red arrow heads depict MCs, T designates thrombus. Yellow arrows show boarder between thrombus and the vessel wall.



Online Figure IV. MC releasate induces ICAM-1 expression on HUVEC in vitro. In vitro differentiated MCs were stimulated with IgE followed by DNP as described in Methods and their releasate added to HUVEC for 20 h. Then, ICAM-1 expression was determined by FACS. ICAM-1 expression upregulation induced by 10 ng/ml TNF- α was taken as 100%.



Online Figure V. DVT is accompanied by elevated plasma sP-selectin levels in WT but not MC-deficient mice. Blood was taken from retroorbital plexus of WT and Kit^{W-sh} mice (n=5 to 9) prior to surgery and 6 h after IVC stenosis application. Plasma levels of sP-selectin were determined using a commercial kit.



Online Figure VI. Systemic administration of histamine H1 antagonists does not protect against DVT. WT mice were injected pyrilamine maleate (upper row; con, n = 9, pyrilamine, n = 10), Cetirizine (lower row; n = 6 in both groups) or sterile saline as described in Methods, and then subjected to 48 h IVC stenosis. (i) Thrombus weight in mg, (ii) thrombus length in mm, (iii) thrombosis prevalence. Horizontal line in dot plots represents median. NS, non-significant.